



ORIGINAL ARTICLE

## Tempol improves lipid profile and prevents left ventricular hypertrophy in LDL receptor gene knockout (LDLr<sup>-/-</sup>) mice on a high-fat diet



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Received 16 June 2016; accepted 13 February 2017

Available online 18 August 2017

### KEYWORDS

Nitroxides;  
Tempol;  
Dyslipidemia;  
Left ventricular hypertrophy;  
Reactive oxygen species;  
Reactive nitrogen species

### Abstract

**Introduction and Objective:** Dyslipidemia is associated with increased risk of cardiovascular disease and atherosclerosis, and hence with high morbidity and mortality. This study investigated the effects of the nitroxide 4-hydroxy-2,2,6,6-tetramethylpiperidine 1-oxyl (Tempol) on lipid profile and cardiac morphology in low-density lipoprotein (LDL) receptor gene knockout (LDLr<sup>-/-</sup>) mice.

**Methods:** Male LDLr<sup>-/-</sup> mice (three months old, approximately 22 g weight) were divided into the following groups: controls, including (1) standard chow (SC, n=8) and (2) high-fat diet (HFD, n=8); and treatment, including (3) standard chow + Tempol (SC+T, n=8) (30 mg/kg administered by gavage, once daily) and (4) high-fat diet + Tempol (HFD+T, n=8) (30 mg/kg). After 30 days of the diet/treatment, whole blood was collected for analysis of biochemical parameters (total cholesterol, triglycerides [TG], high-density lipoprotein [HDL], LDL, and very low-density lipoprotein [VLDL]). The heart was removed through thoracotomy and histological analysis of the left ventricle was performed.

**Results:** A significant increase in TG, LDL, and VLDL and marked left ventricular hypertrophy (LVH) were demonstrated in the HFD group relative to the SC group (p<0.05), while Tempol treatment (HFD+T group) significantly (p<0.05) prevented increases in the levels of these lipid profile markers and attenuated LVH compared with the HFD group.

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**PALAVRAS-CHAVE**

Nitróxidos;  
Tempol;  
Dislipidemia;  
Hipertrofia  
ventricular esquerda;  
EROs/ERNs

**Conclusion:** In this study, Tempol showed potential for the prevention of events related to serious diseases of the cardiovascular system.

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## Tempol melhora o perfil lipídico e previne a hipertrofia ventricular esquerda em camundongos nocaute para o gene do receptor de LDL (LDL<sup>-/-</sup>) sob uma dieta hiperlipídica

**Resumo**

**Introdução e objetivo:** A dislipidemia está associada com aumento do risco para as doenças cardiovasculares e aterosclerose, refletindo na alta morbidade e mortalidade associadas. Este estudo investigou os efeitos do nitróxido 4-Hydroxy-2,2,6,6-tetramethylpiperidine 1-oxyl (tempol) sobre o perfil lipídico e a morfologia cardíaca em camundongos nocaute para o gene do receptor da lipoproteína de baixa densidade (LDLR KO ou LDL<sup>-/-</sup>).

**Métodos:** Camundongos machos (três meses de idade, pesando aproximadamente 22 g) foram divididos nos seguintes grupos: grupos controle: (1) ração padrão ([RP] n=8) = camundongos LDL<sup>-/-</sup> + dieta padrão; (2) dieta rica em lipídios ([DRL] n=8) = camundongos LDL<sup>-/-</sup> + DRL; e grupos tratados: (3) RP + tempol (RP + T, n=8) = camundongos LDL<sup>-/-</sup> + dieta padrão + tempol (30 mg/kg, administrado por gavagem, uma vez por dia); (4) DRL + tempol (DRL + T, n=8) = camundongos LDL<sup>-/-</sup> + DRL + tempol (30 mg/kg). Após 30 dias de dieta/tratamento, o sangue total foi obtido para análise dos parâmetros bioquímicos (colesterol total [CT], triglicerídeos [TG], HDL, LDL e VLDL) e, através de uma toracotomia, o coração foi removido e uma análise histológica do ventrículo esquerdo foi realizada.

**Resultados:** Foi demonstrado um aumento significativo dos níveis de TG, LDL e VLDL, bem como uma considerável hipertrofia ventricular esquerda (HVE), no grupo DRL em comparação com o grupo RP (p<0,05); o tratamento com tempol (grupo DRL + T) preveniu significativamente (p<0,05) o aumento nos níveis destes marcadores de perfil lipídico e atenuou a HVE, em comparação com o grupo DRL.

**Conclusão:** Tempol apresentou potencial para a prevenção de eventos que podem levar a graves doenças do sistema cardiovascular.

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**Introduction**

Cardiovascular disease is the leading cause of morbidity and mortality and is responsible for approximately 30% of all deaths, claiming approximately 17 million lives per year worldwide in 2012.<sup>1-3</sup> Furthermore, many studies have firmly established the relationship between cardiovascular disease and metabolic disorders and underlying conditions such as dyslipidemia, diabetes, and hypertension.<sup>4-8</sup>

Dyslipidemia and associated atherosclerotic/cardiovascular events can present with intense inflammation and increased production of reactive oxygen/nitrogen species (ROS/RNS) from mitochondrial oxidative stress and/or the NADPH oxidase complex, which can cause oxidative modification of LDL, thus amplifying the inflammatory potential (i.e., recruitment of phagocytes and activation of the neutrophil oxidase [Nox]-2 system) and proatherogenic events. Moreover, uncontrolled dyslipidemia can have serious consequences for the cardiovascular system, resulting in morphological changes (left

ventricular hypertrophy [LVH]), dysfunction, and even heart failure.<sup>9-16</sup>

Regulation of lipid metabolism is an important target for therapeutic intervention in dyslipidemic processes to prevent or reduce the risk or severity of cardiovascular disease, and appropriate intervention can have an impact on its clinical course. However, due to the high cost, prolonged use, and especially the adverse effects associated with some lipid-lowering drugs, a drug to control dyslipidemia that presents fewer side effects and a better cost/benefit ratio is highly desirable.<sup>17-19</sup>

Studies have explored other compounds with antioxidant properties in the prevention of cardiovascular disease.<sup>10,20</sup> Over the last few decades, nitroxides have been widely investigated because of their antioxidant capabilities.<sup>20-22</sup> Among them, 4-hydroxy-2,2,6,6-tetramethylpiperidine 1-oxyl (Tempol) is a superoxide dismutase (SOD) mimetic that shows a good partition coefficient, interacts with a broad spectrum of oxidants produced in the human body, and is able to break the chain of redox reactions.<sup>21,23,24</sup>

It has been shown that Tempol has radioprotective, chemopreventive, hypoglycemic, antihypertensive, antineoplastic, and cardioprotective effects, as well as protecting against ischemia-reperfusion injury. It also prevents obesity and neurodegenerative diseases and attenuates renal dysfunction and oxidative stress-induced injury. These biological effects derive at least partially from the ability of this nitroxide to scavenge ROS/RNS, as shown in several studies in which it has been proposed that Tempol can alleviate inflammatory diseases and reduce formation of extracellular traps (ETs) through its action on these oxidants.<sup>22,23,25</sup> However, the beneficial anti-dyslipidemic effects of Tempol and its impact on cardiovascular events remain uncertain, and it is important to understand these actions by exploring different experimental animal models to find new therapeutic options. Therefore, the aim of this study was to assess the effects of Tempol on dyslipidemia and LVH in the well-established animal model of LDLr<sup>-/-</sup> mice.

## Methods

### Ethics statement

All animal experiments were carried out in strict accordance with the recommendations of the Guide for the Care and Use of Laboratory Animals (National Institutes of Health, Washington DC: The National Academy Press, 2011). This study was approved by the ethics committee on the use of animals (CEUA) of the University José do Rosário Vellano (UNIFENAS), approval number 01 A/2015.

### Animals and experimental design

In this study, 40 three-month-old male mice, homozygous for the absence of the LDL receptor gene (LDLr<sup>-/-</sup>), background C57BL6, acquired from the Jackson Laboratory, Bar Harbor, Maine, USA and weighing approximately 22 g were used. These animals were supplied by the UNIFENAS breeding colony and were housed at a controlled temperature (25±1 °C) in a light-controlled room with a 12-h light/dark cycle. After acclimation, the mice were randomly and equally divided into five experimental groups of eight animals per group (n=8), constituted as follows.

### Control groups

The standard chow (SC) group were fed a standard chow (Nuvital<sup>®</sup>, Nuvilab, Colombo, Brazil) for 30 days, and the high-fat diet (HFD) group were fed a high-fat diet (20% total fat, 1.25% cholesterol and 0.5% cholic acid; total 2.89 kcal/g; Instituto Tecnológico de Alimentos, Campinas, SP, Brazil) for 30 days.

### Treatment groups

The standard chow + Tempol (SC+T) group were fed a standard diet (Nuvital<sup>®</sup>) and treated with Tempol (97.0%, Sigma-Aldrich, St. Louis, MO, USA) for 30 days at a dose of 30 mg/kg administered by gavage once daily, and the

high-fat diet + Tempol (HFD+T) group were fed a high-fat diet (20% total fat, 1.25% cholesterol, and 0.5% cholic acid) and treated with Tempol for 30 days at a dose of 30 mg/kg administered by gavage once daily. Additionally, another group (HFD+S) were fed a high-fat diet (20% total fat, 1.25% cholesterol, and 0.5% cholic acid) and treated with simvastatin (Medley, SP, Brazil) at a dose of 20 mg/kg administered by gavage once daily. All animals were fed their respective diets and received water ad libitum.

### Biological samples

After 30 days, mice were maintained on a fasting diet for 12 hours and then anesthetized by intramuscular injection of ketamine (40 mg/kg, Bayer AG and Parke-Davis<sup>®</sup>, Berlin – Bayer, Leverkusen, Germany) and xylazine (6 mg/kg, Bayer AG and Parke-Davis<sup>®</sup>, Berlin – Bayer, Leverkusen, Germany). Absence of the neuromuscular reflex was used to verify the anesthetic effect. Blood was collected via retro-orbital puncture (800 µl) using heparinized capillary tubes. After euthanasia and thoracotomy, 6 ml of 1.34 mM KCl was injected into the hearts through the left ventricle, and the organ was removed.<sup>26</sup>

### Analysis of lipid profiles

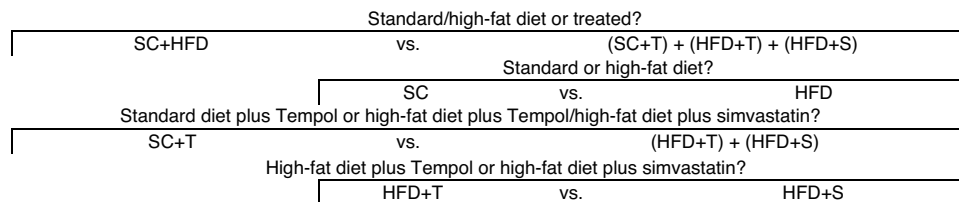
Biochemical markers were assessed by standard methods using commercially available kits (Labtest, MG, Brazil). Plasma levels of total cholesterol (TC, Liquiform kit) and fractions (triglycerides [TG, GPO-ANA kit], high-density lipoprotein [HDL, cholesterol HDL kit], and low-density lipoprotein [LDL, cholesterol LDL kit]) were determined by the endpoint colorimetric method (absorbance values readable spectrophotometrically at 500 or 540 nm). The level of very low-density lipoprotein (VLDL) was determined as previously described.<sup>27</sup>

### Histological analysis of the heart

Briefly, as previously described,<sup>26</sup> the mouse hearts were dissected and the left ventricles were fixed in 10% neutral-buffered formalin for 48 h, and the fixed specimens were processed by a conventional paraffin-embedding technique for histological serial sections of 3-µm thickness. The serial sections were collected from the same plane, deposited on slides, and stained with hematoxylin and eosin for morphological analysis (Nikon optical microscope, TNB-04T-PL, magnification 40× or 100×). Measurement of left ventricular thickness followed standard criteria, using LGMC-image software, version 1.0. All histological analyses were performed by a single examiner using the double-blind method.

### Statistical analysis

The effect of the intervention in the treatment groups was assessed with respect to TG, TC, VLDL, LDL, and HDL. Analysis of variance (ANOVA) was used to determine significant differences between the control and treated groups. Based on the results of the ANOVA/F test, orthogonal contrasts



**Figure 1** Orthogonal contrasts performed. HFD: high-fat diet; HFD+T: high-fat diet + Tempol; HFD+S: high-fat diet + simvastatin; SC: standard chow; SC+T: standard chow + Tempol.

(comparisons) between the variables were performed, as shown in [Figure 1](#).

Before analysis of the data by ANOVA, the Shapiro-Wilk test ( $\alpha=5\%$ ) was performed. This showed that the data had a normal distribution under the null hypothesis of normality for all groups regarding the variables ( $p>0.05$ ). For comparisons of body weight, lipid profile markers and LVH, the mean values  $\pm$  standard error of the mean (SEM) or standard deviation (SD) of at least three experiments are shown, and the variables were analyzed by ANOVA, followed by Tukey's, Scott-Knott's, and Bonferroni's tests for multiple comparisons of the means ( $\alpha=5\%$ ). Additionally, interactions between the groups and times (0 and 30 days) were considered, with body weight being assessed at 0 and 30 days. The main effects were also assessed separately for groups and times in terms of body weight. Sisvar (Lavras, MG, Brazil, 2008) and BioEstat 5.0 (Belém, Pará, Brazil, 2007) were used for the statistical analysis.

## Results

### Effects of Tempol on the body weights of LDLr<sup>-/-</sup> mice

Results of the comparisons of body weights according to the times at the beginning and end of the study period are shown in [Table 1](#). No significant differences were observed between the groups over time (at 0 and after 30 days)

( $p>0.05$ ) ([Table 1](#), a). [Table 1](#) also shows the overall mean weight (all groups) at 0 and 30 days, which was observed to increase after 30 days of the experiment ([Table 1](#), b). In contrast, [Table 2](#) shows that, in general, there was a decrease in mean body weight in the treated groups (SC+T, HFD+T, and HFD+S) compared to the control groups (SC and HFD vs. others) ( $p=0.000$ ); however, there was no significant difference among the controls (SC vs. HFD). The treated groups did not differ statistically ( $p=0.886$ ).

### Effects of Tempol on triglycerides, total cholesterol, very low-density lipoprotein, low-density lipoprotein, and high-density lipoprotein in LDLr<sup>-/-</sup> mice

Treatment with Tempol in the HFD+T group had no effect on TC levels ([Figure 2](#)), but prevented increases in plasma levels of TG, VLDL, and LDL compared with the control HFD group ([Figures 3–5](#)). Regarding the orthogonal contrasts assessing the intervention for the treated groups vs. control groups (all untreated groups or baseline groups), [Table 2](#) shows that the treated groups (SC+T, HFD+T, and HFD+S) differed from the untreated groups (SC and HFD) ( $p=0.000$ ) in terms of TC; however, this variable did not differ significantly when the groups were compared to each other ([Figure 2](#)). [Table 2](#) also shows that TG levels fell in the treated groups compared with the untreated groups ( $p=0.000$ ), a similar result to that presented in [Figure 2](#). Regarding LDL ([Table 2](#)), the groups

**Table 1** Means of body weights assessed in the study groups compared at 0 and 30 days.

a.	Times <sup>b</sup>	
	0 days	30 days
Experimental groups <sup>a</sup>	Weight (g)	
SC	23.47 <sup>a A</sup>	22.86 <sup>a A</sup>
HFD	21.77 <sup>a A</sup>	24.43 <sup>a A</sup>
SC+T	19.19 <sup>a A</sup>	21.07 <sup>a A</sup>
HFD+T	19.68 <sup>a A</sup>	20.59 <sup>a A</sup>
HFD+S	20.10 <sup>a A</sup>	20.16 <sup>a A</sup>
b. Times (days)	Means <sup>c</sup>	
0	20.44 <sup>a</sup>	
30	21.84 <sup>b</sup>	

HFD: high-fat diet; HFD+T: high-fat diet + Tempol; HFD+S: high-fat diet + simvastatin; SC: standard chow; SC+T: standard chow + Tempol.

<sup>a</sup> Means followed by the same superscript uppercase letter (row) do not differ by Tukey's test ( $\alpha=5\%$ ).

<sup>b</sup> Means followed by the same superscript lowercase letter (column) do not differ by Tukey's test ( $\alpha=5\%$ ).

<sup>c</sup> Means followed by the same letter do not differ statistically by the Student's t test ( $\alpha=5\%$ ).

**Table 2** p-values, estimates, and coefficients for the variables analyzed according to the orthogonal contrasts shown in Figure 1.

Groups	SC	HFD	SC+T	HFD+T	HFD+S
Contrasts (coefficients)					
SC and HFD vs. others	3	3	-2	-2	-2
SC vs. HFD	1	-1	0	0	0
SC+T vs. HFD+T and HFD+S	0	0	2	-1	-1
HFD+T vs. HFD+S	0	0	0	1	-1
Body weight			Total cholesterol		
Contrasts			Contrasts		
Groups	Estimates	p	Groups	Estimates	p
SC and HFD vs. others	2.52	0.000 <sup>b</sup>	SC and HFD vs. others	131.83	0.000 <sup>b</sup>
SC vs. HFD	-0.94	0.077 <sup>c</sup>	SC vs. HFD	157.42	0.001 <sup>b</sup>
SC+T vs. HFD+T and HFD+S	-0.06	0.886 <sup>c</sup>	SC+T vs. HFD+T and HFD+S	73.90	0.058 <sup>c</sup>
HFD+T vs. HFD+S	0.00	0.992 <sup>c</sup>	HFD+T vs. HFD+S	-25.25	0.297 <sup>c</sup>
Triglycerides			LDL		
Contrasts			Contrasts		
Groups	Estimates	p	Groups	Estimates	p
SC and HFD vs. others	164.40	0.000 <sup>b</sup>	SC and HFD vs. others	70.54	0.007 <sup>b</sup>
SC vs. HFD	162.87	0.016 <sup>a</sup>	SC vs. HFD	216.48	0.000 <sup>b</sup>
SC+T vs. HFD+T and HFD+S	144.48	0.014 <sup>a</sup>	SC+T vs. HFD+T and HFD+S	117.18	0.001 <sup>b</sup>
HFD+T vs. HFD+S	-25.25	0.696 <sup>c</sup>	HFD+T vs. HFD+S	-36.82	0.332 <sup>c</sup>
VLDL			HDL		
Contrasts			Contrasts		
Groups	Estimates	p	Groups	Estimates	p
SC and HFD vs. others	25.94	0.000 <sup>b</sup>	SC and HFD vs. others	0.077	0.990 <sup>c</sup>
SC vs. HFD	60.01	0.000 <sup>b</sup>	SC vs. HFD	-22.75	0.221 <sup>c</sup>
SC+T vs. HFD+T and HFD+S	37.50	0.000 <sup>b</sup>	SC+T vs. HFD+T and HFD+S	5.01	0.535 <sup>c</sup>
HFD+T vs. HFD+S	-18.21	0.083 <sup>c</sup>	HFD+T vs. HFD+S	-25.25	0.957 <sup>c</sup>
LVH			Contrasts		
Groups	Estimates	p			
SC and HFD vs. others			0.456		0.003 <sup>b</sup>
SC vs. HFD			0.812		0.001 <sup>b</sup>
SC+T vs. HFD+T and HFD+S			0.783		0.782 <sup>c</sup>
HFD+T vs. HFD+S			0.461		0.036 <sup>a</sup>

HDL: high-density lipoprotein; HFD: high-fat diet; HFD+T: high-fat diet + Tempol; HFD+S: high-fat diet + simvastatin LDL: low-density lipoprotein; LVH: left ventricular hypertrophy; SC: standard chow; SC+T: standard chow + Tempol.

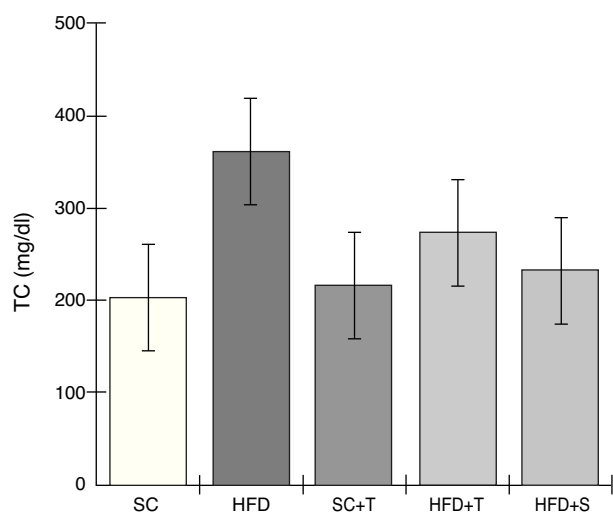
<sup>a</sup> Significant at a nominal level of 5% (p<0.05).  
<sup>b</sup> Significant at a nominal level of 1% (p<0.01).  
<sup>c</sup> Not significant at a nominal level of 5% (p>0.05).

differed (p=0.000); the treated groups (SC+T, HFD+T, and HFD+S) presented lower mean LDL values than in the control groups (SC and HFD) (Table 2, Figure 4). No significant difference (p=0.332) was observed between the HFD+T and HFD+S groups (Figure 4). Table 2 shows that the treated groups (SC+T, HFD+T, and HFD+S) presented a decrease in VLDL compared to the control groups (SC and HFD) (p=0.000). There was no significant difference between the HFD+T and HFD+S groups (p=0.083) (Figure 5). No significant differences were

observed in HDL between the treated and untreated groups (Table 2, p=0.1840) and among all groups (Figure 6).

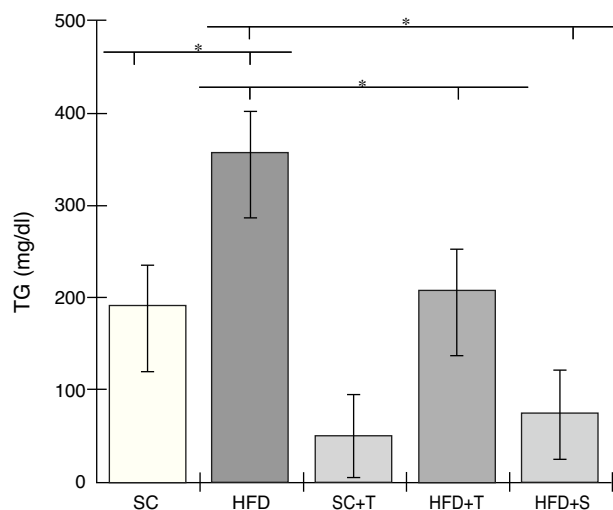
### Effects of Tempol on cardiac damage and left ventricular hypertrophy in LDLr<sup>-/-</sup> mice

Regarding cardiac remodeling, representative histological images of the experimental groups are shown in Figure 6.

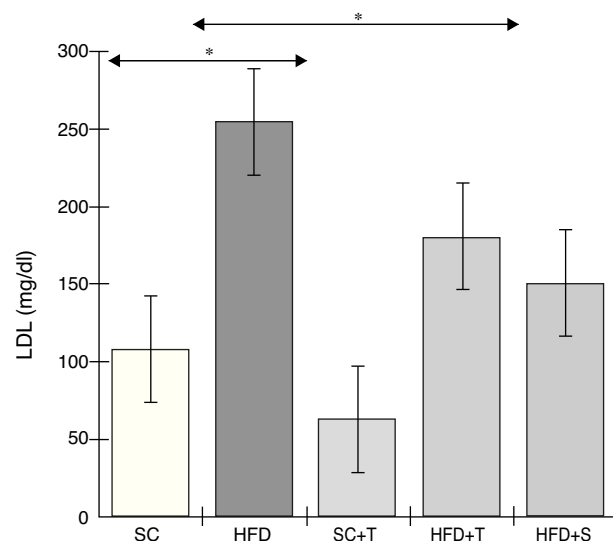


**Figure 2** Blood levels of total cholesterol in the experimental groups fed a standard diet and a high-fat diet. HFD: high-fat diet; HFD+T: high-fat diet + Tempol; HFD+S: high-fat diet + simvastatin; SC: standard chow; SC+T: standard chow + Tempol; TC: total cholesterol. Values are mean  $\pm$  standard error of the mean;  $\alpha=0.05$ ; \* $p<0.05$ .

The data show a correlation between morphology and left ventricular thickness. The mean thicknesses (in  $\mu\text{m}$ ) in the five experimental groups are shown in Figure 7. There was no difference between the SC+T and SC groups ( $p=\text{NS}$ ). Mean thickness was 0.6  $\mu\text{m}$  less in the HFD+T group ( $p<0.05$ ) than in the HFD group, and was 1.1  $\mu\text{m}$  less in the HFD+S group ( $p<0.05$ ) than in the HFD group and 0.5  $\mu\text{m}$  less than in the HFD+T group. The difference between HFD+S and HFD+T was not significant ( $p>0.05$ ) (Figure 8). Table 2 shows that, in general, the treated groups (SC+T, HFD+T, and HFD+S) presented less LVH than the control groups (SC and HFD)



**Figure 3** Blood levels of triglycerides in the experimental groups fed a standard diet and a high-fat diet. HFD: high-fat diet; HFD+T: high-fat diet + Tempol; HFD+S: high-fat diet + simvastatin; SC: standard chow; SC+T: standard chow + Tempol; TG: triglycerides. Values are mean  $\pm$  standard error of the mean;  $\alpha=0.05$ ; \* $p<0.05$ .

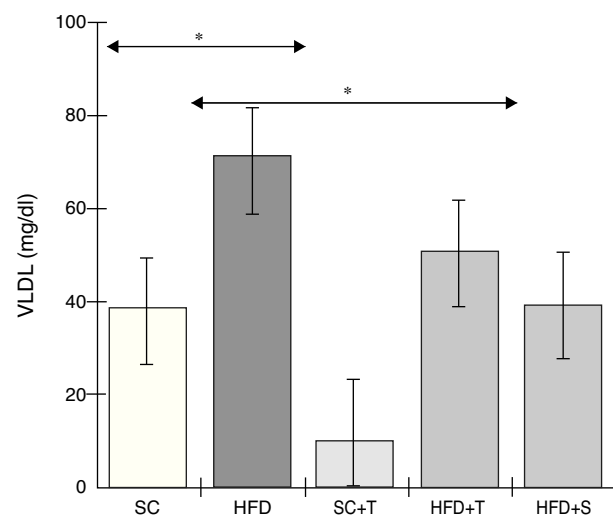


**Figure 4** Blood levels of low-density lipoprotein in the experimental groups fed a standard diet and a high-fat diet. HFD: high-fat diet; HFD+T: high-fat diet + Tempol; HFD+S: high-fat diet + simvastatin; LDL: low-density lipoprotein; SC: standard chow; SC+T: standard chow + Tempol. Values are mean  $\pm$  standard error of the mean;  $\alpha=0.05$ ; \* $p<0.05$ .

( $p=0.003$ ). Additionally, based on analyses between the individual groups (Figure 7), the control groups (baseline, SC and HFD) were significantly different from each other ( $p=0.001$ ).

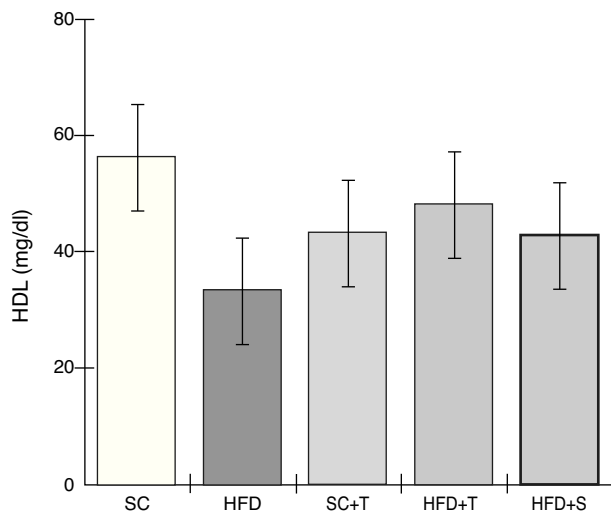
## Discussion

In this study, we used a well-established animal model of LDL receptor knockout (LDLR<sup>-/-</sup>) mice, whose genetic background combined with a high-fat diet (environmental factor)

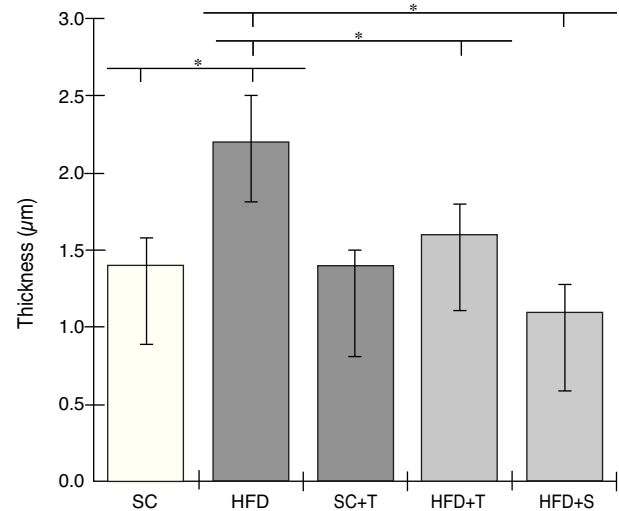


**Figure 5** Blood levels of very low-density lipoprotein in the experimental groups fed a standard diet and a high-fat diet. HFD: high-fat diet; HFD+T: high-fat diet + Tempol; HFD+S: high-fat diet + simvastatin; SC: standard chow; SC+T: standard chow + Tempol; VLDL: very low-density lipoprotein. Values are mean  $\pm$  standard error of the mean;  $\alpha=0.05$ ; \* $p<0.05$ .





**Figure 6** Blood levels of high-density lipoprotein in the experimental groups fed a standard diet and a high-fat diet. HFD: high-fat diet; HFD+T: high-fat diet + Tempol; HFD+S: high-fat diet + simvastatin; HDL: high-density lipoprotein; SC: standard chow; SC+T: standard chow + Tempol. Values are mean  $\pm$  standard error of the mean;  $\alpha=0.05$ ; \* $p<0.05$ .



**Figure 8** Mean left ventricular thickness in the study groups. HFD: high-fat diet; HFD+T: high-fat diet + Tempol; SC: standard chow; SC+T: standard chow + Tempol; HFD+S: high-fat diet + simvastatin. Values are mean  $\pm$  standard deviation;  $\alpha=0.05$ ; \* $p<0.05$ .

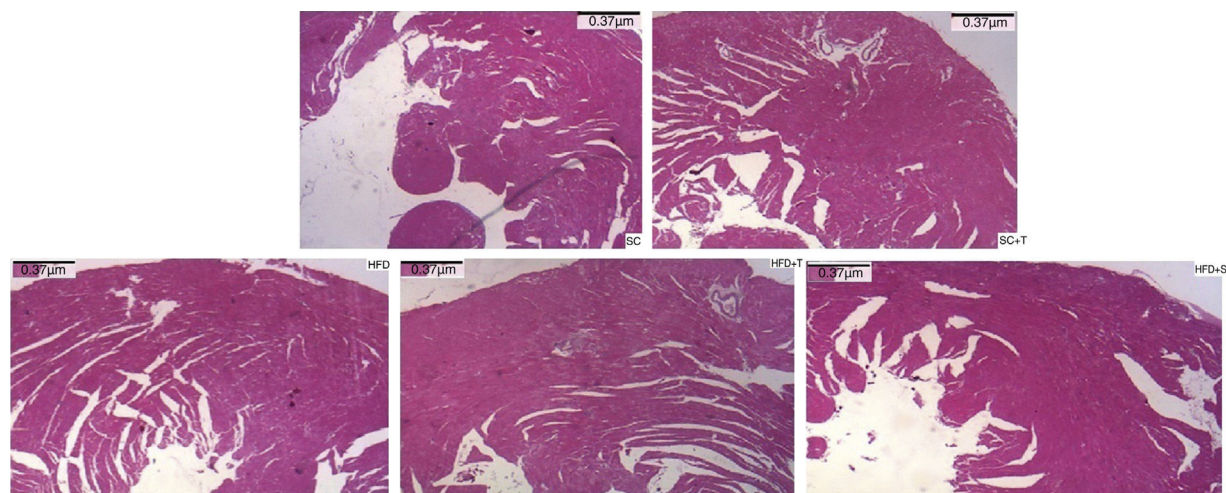
is likely to lead to the development of severe dyslipidemia, and hence a high risk of cardiovascular disease. During the study, LDLr<sup>-/-</sup> mice were fed a standard or a high-fat diet for 30 days. Administration of Tempol (30 mg/kg once daily for 30 days) in mice on a high-fat diet showed a strong protective effect in controlling dyslipidemia and preventing damage to heart tissue (mitigating LVH).

LDLr<sup>-/-</sup> mice on a high-fat diet treated with Tempol showed a marked decrease in TG, LDL, and VLDL levels. Among the lipoproteins, LDL has been identified as one of the most important constituents of atheroma. As expected, LDLr<sup>-/-</sup> mice fed a high-fat diet presented a marked increase in LDL levels, and Tempol treatment decreased levels of this lipoprotein. Furthermore, increased TG levels have recently been shown to be associated with low HDL cholesterol levels,

and hypertriglyceridemia is also one of the 'deadly quartet,' along with abdominal obesity, hypertension, and glucose intolerance.<sup>7,28,29</sup>

In our study, and in agreement with Kim et al.,<sup>11</sup> LDLr<sup>-/-</sup> mice developed dyslipidemia with increases in TG, VLDL, and LDL (on a high-fat diet), and Tempol attenuated this condition; these results are similar to those found by Kim et al., in which Tempol treatment also prevented increases in these biomarkers of dyslipidemia.

Lipid disorders are among the most important risk factors for atherosclerotic cardiovascular disease, together with other chronic degenerative diseases with a prolonged natural history such as hypertension, obesity, and diabetes.<sup>4-7</sup> These diseases have a complex relationship with one another; lifestyle and genetic inheritance, among



**Figure 7** Morphological analysis of the left ventricle in LDLr<sup>-/-</sup> mice fed a standard diet (above) and a high-fat diet (below). HFD: high-fat diet; HFD+T: high-fat diet + Tempol; SC: standard chow; SC+T: standard chow + Tempol; HFD+S: high-fat diet + simvastatin.

other factors, are common to their etiologies. Treatment of dyslipidemia has the fundamental purpose of primary and secondary prevention of coronary artery disease, cerebrovascular disease, and peripheral arterial disease, and may also lead to the regression of xanthomas and reduce the risk of acute pancreatitis.<sup>30–32</sup>

Unlike LDL, HDL functions primarily in reverse cholesterol transport, reducing the formation of atherosclerotic plaque.<sup>33</sup> In this study, HDL and TC levels in LDLr-/- mice did not differ significantly among the five groups evaluated. By contrast, the plasma HDL levels found by Kim et al.<sup>11</sup> demonstrated that treatment with Tempol of apoE-/- mice maintained on a high-fat diet promoted increases in HDL levels, but these differences may be due to the characteristics of the different animal models used.<sup>34</sup>

Even on a normal diet, LDLr-/- mice may slowly develop dyslipidemia and atherosclerosis over time, and this process is accelerated with a high-fat diet.<sup>34</sup> LDLr-/- mice develop moderate hypercholesterolemia (TC ~250 mg/dl) when fed a standard diet and severe hypercholesterolemia when fed a high-fat diet. However, no differences between the groups were found regarding TC and HDL in this study; moreover, a closer relationship between these biomarkers was found. Furthermore, LDLr-/- mice normally become obese only when fed a high-fat diet with a fat content of more than 20%; thus, body weights differed only slightly among the groups, without statistical significance (Table 1) since only up to 20% total fat was used in this study.<sup>34</sup>

Simvastatin was used in this study as a control since this cholesterol-lowering agent is widely used preventively or as an adjuvant to correct lipid metabolism in patients with a predisposition to cardiovascular disease or primary hypercholesterolemia. Simvastatin is a prodrug that acts on the enzyme HMG-CoA reductase, preventing cholesterol synthesis and decreasing LDL, VLDL and TG.<sup>19,35</sup> This statin, at a dose of 20 mg/kg, had significant effects on lipid profile markers and LVH, and its effects on TG, LDL, VLDL, and LVH were statistically similar to those caused by 30 mg/kg Tempol.

In LVH, increases in myocyte volume, coronary artery wall thickness, capillary rarefaction and extracellular fibrosis, and changes in energy metabolism, intracellular calcium, and myocardial contractility and relaxation are normally observed.<sup>13,14</sup> LDLr-/- mice fed a high-fat diet for approximately 14 days develop atherosclerosis and an increased predisposition to cardiac damage, including LVH, with a mean 30% increase in cardiomyocyte diameter.<sup>26</sup> In our study, morphological changes were observed in the untreated group of LDLr-/- mice fed a high-fat diet compared with those fed a standard diet, while treatment of these mice with Tempol attenuated these alterations, preventing LVH.

Regarding the underlying mechanisms, studies have shown that superoxide ( $O_2^{\cdot-}$ ) production in LDLr-/- mice fed a high-fat diet is significantly higher than in LDLr-/- mice fed standard chow.<sup>36,37</sup> Moreover, oxidative stress in these mice decreases nitric oxide (\*NO) and increases LDL oxidation. Thus, it is possible that antioxidant treatment reduces cardiac oxidative stress and prevents LVH by mitigating pathogenic mechanisms such as oxidative stress-mediated fibrosis.<sup>38</sup>

Ulasova et al. showed that quercetin, an antioxidant, can prevent LVH in ApoE-/- hypercholesterolemic mice,<sup>39</sup> and other studies have shown that Tempol treatment can inhibit hypertension-induced oxidative stress-related LVH. In addition, Tempol (3 mmol) added to the drinking water of an experimental model of Dahl salt-sensitive rats for 10 weeks normalized LVH and reduced the cardiac expression of p22phox and Nox-2, mitochondrial uncoupling protein 2 and related oxidative stress. Thus, inhibition of cardiac ROS by Tempol prevented the cardiac fibrosis, remodeling and defective relaxation that underlie diastolic heart failure.<sup>40–42</sup>

The accumulated evidence suggests that Tempol can exert positive effects on dyslipidemia and prevent cardiac damage through multiple mechanisms; furthermore, a pleiotropic action is more plausible than a single mechanism in explaining the restoration of \*NO and nitric oxide synthetase (NOS) (\*NO is an important regulator of cardiac remodeling and is recognized as an anti-hypertrophic mediator).<sup>43,44</sup>

Furthermore, in this context, the action of Tempol in decreasing ROS is also relevant, since an increase in ROS in cardiomyocytes can activate the MAPK pathway, which has an important role in cardiac hypertrophy, and redox imbalance can also decrease the bioavailability of \*NO.<sup>43</sup> Other ROS/RNS scavengers have been described as possible options for controlling atherosclerotic events and/or cardiovascular disease-related oxidative stress, since decreased antioxidant (i.e. SOD) activity and increased ROS generation have been described in such conditions, as in LVH.<sup>45–50</sup>

Several studies using other experimental models have reported the effects of Tempol on dyslipidemia, atherosclerotic events, and the cardiovascular system. The cardioprotective effect of Tempol demonstrated in this study using an LDLr-/- mice model is consistent with the findings of Zhu et al.,<sup>51</sup> who demonstrated the action of Tempol (500  $\mu$ M) on  $O_2^{\cdot-}$  in rat aortas, with a protective effect on the cardiovascular system by increasing the release of NO in the endothelium and causing vascular relaxation in aortas. Furthermore, the authors demonstrated that chronic treatment with Tempol (1 mM) added to the mice's drinking water restored the release of NO and aortic relaxation in rats.

Similarly to our findings, in an experimental model of rats with caerulein-induced pancreatitis, Marciniak et al.<sup>32</sup> demonstrated that Tempol significantly decreased myocardial damage, mainly by attenuating oxidative stress-induced damage. Furthermore, as cited above, Kim et al.,<sup>11</sup> using an experimental ApoE-/- mouse model, showed that Tempol (10 mg/g added to feed) can improve lipid profile, reduce the formation of pro-inflammatory cytokines and markers such as monocyte chemoattractant protein and myeloperoxidase, helping reduce atherosclerotic plaque area and risk of myocardial damage.

Our study has potential limitations. Assessment of oxidative damage by measuring oxidative stress indices would have helped to demonstrate the role of oxidants in the events described herein. Measurement of other biochemical parameters such as blood glucose, interleukin-6, monocyte-chemoattractant protein, myeloperoxidase, and serum amyloid A could also have supplemented these findings and established a stronger link between dyslipidemia, the genesis



of atherosclerotic events, and subsequent cardiovascular complications.

Regarding the Tempol dose used in this study (30 mg/kg/day), other studies have reported the use of this nitroxide in animal models at doses of approximately 30 mg/kg or concentrations from 100 to 1000  $\mu\text{M}/\text{kg}$ , once daily.<sup>11,32,51</sup> These doses and concentrations do not display in-vivo toxicity (even doses of 300 mg/kg/day or 87  $\mu\text{M}/\text{kg}$  have no toxic effects in vivo),<sup>23,52</sup> are compatible with an EC50 for inhibiting the oxidative burst in vitro (50-400  $\mu\text{M}$ ),<sup>53</sup> and concentrations up to 1000  $\mu\text{M}$  appear to be safe, i.e., it does not show in-vitro pro-oxidant effects,<sup>54,55</sup> which we consider may be a significant dose-limiting side effect in the systemic use of this nitroxide.

In addition to Tempol's previously mentioned possible side effects, caution should be exercised with nitroxides and other antioxidants because of the reductive stress that can occur with excessive use of this and other antioxidants, which may have short-, medium- and long-term effects. It has been reported that antioxidants influence the immune response (acting mainly by inhibiting the oxidative burst) and the onset of chronic diseases, such as cancer and cardiovascular disease, including cardiomyopathy.<sup>56</sup>

In summary, our study shows that the nitroxide Tempol is able to improve lipid profile and attenuate LVH in LDLr<sup>-/-</sup> mice fed a high-fat diet. These data suggest that this antioxidant can be a potent ally in preventing events related to cardiovascular disease.

## Ethical disclosures

**Protection of human and animal subjects.** The authors declare that the procedures followed were in accordance with the regulations of the relevant clinical research ethics committee and with those of the Code of Ethics of the World Medical Association (Declaration of Helsinki).

**Confidentiality of data.** The authors declare that no patient data appear in this article.

**Right to privacy and informed consent.** The authors declare that no patient data appear in this article.

## Conflicts of interest

The authors have no conflicts of interest to declare.

## Acknowledgments

This research was supported by a grant from Fundação de Amparo à Pesquisa do Estado de Minas Gerais.

## References

- Bhatnagar P, Wickramasinghe K, Williams J, et al. The epidemiology of cardiovascular disease in the UK 2014. *Heart*. 2015;101:1182–9.
- Rosenbaugh EG, Savalia KK, Manickam DS, et al. Antioxidant-based therapies for angiotensin II-associated cardiovascular diseases. *Am J Physiol Regul Integr Comp Physiol*. 2013;304:R917–28.
- World Health Organization. The top 10 causes of death: major causes of death. Available at: <http://www.who.int/mediacentre/factsheets/fs310/en/index2.html> [accessed November 2016].
- Castelli WP. Epidemiology of coronary heart disease: the Framingham Study. *Am J Med*. 1984;27:4–12.
- Zimmet ZP. Obesity, hypertension, carbohydrate disorders and risk of chronic diseases. *Med J Aust*. 1986;145:256–62.
- Levy D. Cardiovascular risks: new insights from Framingham. *Am Heart J*. 1988;116:266–72.
- Kaplan NM. The deadly quartet. *Arch Intern Med*. 1989;149:1514–20.
- Eckel R, Krauss R. American Heart Association call to action: obesity as a major risk factor for coronary heart disease. *Circulation*. 1998;97:2099–100.
- Yoshida WB. Radicais livres na síndrome da isquemia e reperfusão. *Cir Vasc Angiol*. 1996;12:82–95.
- Cook NR, Albert CM, Gaziano JM, et al. A randomized factorial trial of vitamins C and E and beta-carotene in the secondary prevention of cardiovascular events in women: results from the Women's Antioxidant Cardiovascular Study. *Arch Intern Med*. 2007;67:1610–8.
- Kim CH, Mitchell JB, Bursill CA, et al. The nitroxide radical TEMPOL prevents obesity, hyperlipidaemia, elevation of inflammatory cytokines, and modulates atherosclerotic plaque composition in apoE<sup>-/-</sup> mice. *Atherosclerosis*. 2015;240:234–41.
- Ong S-B, Hall AR, Hausenloy DJ. Mitochondrial dynamics in cardiovascular health and disease. *Antioxid Redox Signal*. 2013;19:400–14.
- Gradman AH, Alfayoumi F. From left ventricular hypertrophy to congestive heart failure: management of hypertensive heart disease. *Prog Cardiovasc Dis*. 2006;48:326–41.
- Verdecchia P, Angeli F, Achilli P, et al. Echocardiographic left ventricular hypertrophy in hypertension: marker for future events or mediator of events? *Curr Opin Cardiol*. 2007;22:329–34.
- Lambeth JD. Nox enzymes, ROS, and chronic disease: an example of antagonistic pleiotropy. *Free Radic Biol Med*. 2007;43:332–47.
- Martínez-Revelles S, Avendaño MS, García-Redondo AB, et al. Reciprocal relationship between reactive oxygen species and cyclooxygenase-2 and vascular dysfunction in hypertension. *Antioxid Redox Signal*. 2012.
- Schaefer EJ. Lipoproteins nutrition, and heart disease. *Am J Clin Nutr*. 2002;75:191–212.
- Kingsbury KJ, Bondy G. Understanding the essentials of blood lipid metabolism. *Prog Cardiovasc Nurs*. 2003;18:13–8.
- Golomb BA, Evans MA. Statin adverse effects: a review of the literature and evidence for a mitochondrial mechanism. *Am J Cardiovasc Drugs*. 2008;8:373–418.
- Soule BP, Hyodo F, Matsumoto K, et al. The chemistry and biology of nitroxide compounds. *Free Radic Biol Med*. 2007;42:1632–50.
- Israeli A, Patt M, Oron M, et al. Kinetics and mechanism of the comproportionation reaction between oxoammonium cation and hydroxylamine derived from cyclic nitroxides. *Free Radic Biol Med*. 2005;38:317–24.
- Wilcox CS, Pearlman A. Chemistry and antihypertensive effects of Tempol and other nitroxides. *Pharmacol Rev*. 2008;60:418–69.
- Wilcox CS. Effects of Tempol and redox-cycling nitroxides in models of oxidative stress. *Pharmacol Ther*. 2010;126:119–45.
- Samuni AM, Barenholz Y. Site-activity relationship of nitroxide radical's antioxidative effect. *Free Radic Biol Med*. 2003;34:177–85.
- Hosseinzadeh A, Messer PK, Urban CF. Stable redox-cycling nitroxide Tempol inhibits NET formation. *Front Immunol*. 2012;3:391.

26. Garcia JAD, Lima CC, Messora LB, et al. Efeito anti-inflamatório da lipoproteína de alta densidade no sistema cardiovascular de camundongos hiperlipidêmicos. *Rev Port Cardiol*. 2011;30:763–9.
27. Tian J, Pei H, Sanders JM, et al. Hyperlipidemia is a major determinant of neointimal formation in LDL receptor-deficient mice. *Biochem Biophys Res Commun*. 2006;345:1004–9.
28. Carlson LA, Böttiger LE. Serum triglycerides, to or not to be a risk factor for ischaemic heart disease? *Atherosclerosis*. 1981;39:289–91.
29. Assman G, Schulte H. Role of triglycerides in coronary artery disease: lessons from the Prospective Cardiovascular Münster Study. *Am J Cardiol*. 1992;70:10H–3H.
30. Shieh SM, Shen M, Fuh MM, et al. Plasma lipid and lipoprotein concentrations in Chinese males with coronary artery disease, with and without hypertension. *Atherosclerosis*. 1987;67:49–55.
31. La Rosa JL. Cholesterol and cardiovascular disease: how strong is the evidence? *Clin Cardiol*. 1992;15 Suppl. III:2–7.
32. Marciniak A, Walczyna B, Rajtar G, et al. Tempol, a membrane-permeable radical scavenger, exhibits anti-inflammatory and cardioprotective effects in the cerulein-induced pancreatitis rat model. *Oxid Med Cell Longev*. 2016;2016. Article ID 4139851, 7 pp.
33. Arntzenius AC. Regression of atherosclerosis. Benefit can be expected from low LDL-C and high HDL-C levels. *Acta Cardiol*. 1991;4:431–8.
34. Kennedy AJ, Ellacott KLJ, King VL, et al. Mouse models of the metabolic syndrome. *Dis Model Mech*. 2010;3:156–66.
35. Malenovic A, Jancic-stojanovic B, Medenica M, et al. Microemulsion liquid chromatographic screening of simvastatin and its active metabolite in human plasma. *Acta Chromatogr*. 2008;20:595–607.
36. Krieger MH, Santos KFR, Shishido SM, et al. Antiatherogenic effects of S-nitroso-N-acetylcysteine in hypercholesterolemic LDL receptor knockout mice. *Nitric Oxide*. 2006;14:12–20.
37. Wanschel AC, Caceres VM, Moretti AI, et al. Cardioprotective mechanism of S-nitroso-N-acetylcysteine via S-nitrosated betaadrenoceptor-2 in the LDLr<sup>-/-</sup> mice. *Nitric Oxide*. 2014;36:58–66.
38. Rizzi E, Castro MM, Ceron CS, et al. Tempol inhibits TGF- $\beta$  and MMPs upregulation and prevents cardiac hypertensive changes. *Int J Cardiol*. 2013;165:165–73.
39. Ulasova E, Perez J, Hill BJ, et al. Quercetin prevents left ventricular hypertrophy in the Apo E knockout mouse. *Redox Biol*. 2013;1:381–6.
40. Guo P, Nishiyama A, Rahman M, et al. Contribution of reactive oxygen species to the pathogenesis of left ventricular failure in Dahl salt-sensitive hypertensive rats: effects of angiotensin II blockade. *J Hypertens*. 2006;24:1097–104.
41. Matsui H, Ando K, Kawarazaki H, et al. Salt excess causes left ventricular diastolic dysfunction in rats with metabolic disorder. *Hypertension*. 2008;52:287–94.
42. Wang H, Shimosawa T, Matsui H, et al. Paradoxical mineralocorticoid receptor activation and left ventricular diastolic dysfunction under high oxidative stress conditions. *J Hypertens*. 2008;26:1453–62.
43. Garcia JAD, Incerpi EK. Factors and mechanisms involved in left ventricular hypertrophy and the anti-hypertrophic role of nitric oxide. *Arq Bras Cardiol*. 2008;90:409–16.
44. Zarlino JA, Brunt VE, Vallerga AK, et al. Nitroxide pharmaceutical development for age-related degeneration and disease. *Front Genet*. 2015;6:325.
45. Balta N, Stojan I, Petec C, et al. Decreased SOD activity and increased nitrates level in rat heart with left ventricular hypertrophy induced by isoproterenol. *Rom J Physiol*. 1999;36:175–82.
46. Lu Z, Xu X, Hu X, et al. Extracellular superoxide dismutase deficiency exacerbates pressure overload-induced left ventricular hypertrophy and dysfunction. *Hypertension*. 2008;51:19–25.
47. Afanas'ev I. ROS and RNS signaling in heart disorders: could antioxidant treatment be successful? *Oxid Med Cell Longev*. 2011;2011. Article ID 293769, 13 pp.
48. Dickey JS, Gonzalez Y, Aryal B, et al. Mito-tempol and dexrazoxane exhibit cardioprotective and chemotherapeutic effects through specific protein oxidation and autophagy in a syngeneic breast tumor preclinical model. *PLOS ONE*. 2013;8:e70575.
49. van Deel ED, Lu Z, Xu X, et al. Extracellular SOD protects the heart against oxidative stress and hypertrophy after myocardial infarction. *Free Radic Biol Med*. 2008;44:1305–13.
50. Hoffman A, Goldstein S, Samuni A, et al. Effect of nitric oxide and nitroxide SOD-mimic on the recovery of isolated rat heart following ischemia and reperfusion. *Biochem Pharmacol*. 2003;66:1279–86.
51. Zhu J, Drenjancevic-Peric N, McEwen E, et al. Role of superoxide and angiotensin II suppression in salt-induced changes in endothelial Ca<sup>2+</sup> signaling and NO production in rat aorta. *Am J Physiol Heart Circ Physiol*. 2006;291:H929–38.
52. Metz JM, Smith D, Mick R, et al. A phase I study of topical Tempol for the prevention of alopecia induced by whole brain radiotherapy. *Clin Cancer Res*. 2004;10:6411–7.
53. Jackson TC, Mi Z, Jackson EK. Modulation of cyclic AMP production by signal transduction pathways in preglomerular microvessels and microvascular smooth muscle cells. *J Pharmacol Exp Ther*. 2004;310:349–58.
54. Aronovitch Y, Godinger D, Israeli A, et al. Dual activity of nitroxides as pro- and antioxidants: catalysis of copper-mediated DNA breakage and H<sub>2</sub>O<sub>2</sub> dismutation. *Free Radic Biol Med*. 2007;42:1317–25.
55. Głebska J, Skolimowski J, Kudzin Z, et al. Pro-oxidative activity of nitroxides in their reactions with glutathione. *Free Radic Biol Med*. 2003;35:310–6.
56. Brewer AC, Mustafi SB, Murray TVA, et al. Reductive stress linked to small HSPs, G6PD, and Nrf2 pathways in heart disease. *Antioxid Redox Signal*. 2013;18:1114–27.