



Original article

Genetic polymorphism in DNMTs and gastric cancer: A systematic review and meta-analysis



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ABSTRACT

Epigenetics alterations, including aberrant DNA methylation, have been associated with gastric carcinogenesis. Single nucleotide polymorphisms (SNPs) in DNA methyltransferases (DNMTs) may influence protein expression and therefore affect DNA regulation and susceptibility for Gastric Cancer (GC).

We have performed a systematic review and meta-analysis involving 11 studies and a total of 24 SNPs in DNMTs were analyzed. According to literature, only 4 SNPs, DNMT1 rs16999593, DNMT2 rs11254413 and DNMT3A rs7560488 and DNMT3A rs36012910, were associated with GC. DNMT1 rs16999593 and DNMT3A rs7560488C allele and DNMT3A rs36012910 G allele showed an increased risk for GC. On the other hand, DNMT2 rs11254413 G allele presented a protective effect for GC. Additionally, the meta-analysis evaluated the SNPs analyzed in more than one study ($n = 6$). Results revealed that only DNMT1 rs16999593 had a statistically significant association with GC development (OR = 1.31; 95% CI = 1.08–1.60; $p = 0.006$ for TC + CC genotypes).

Our study suggests that DNMT2 rs11254413, DNMT3A rs7560488, DNMT3A rs36012910 and, specially, DNMT1 rs16999593 may have an association with GC development. Nevertheless, further studies are need using different populations to clarify this association with GC risk.

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Introduction

Epigenetics, and more extensively DNA methylation, has been studied in the past decades and associated with several diseases, including cancer. Literature shows that altered epigenetic control of gene expression has an important role in carcinogenesis.¹ DNA methylation is an important epigenetic mark in tumorigenesis, in which a methyl group is added to the 5' position of the cytosine residue in a CpG dinucleotide (CpG islands, CGI), in the 5'-flanking promoter's genes.² Aberrant methylation of promoter's genes is an important hallmark of cancer cells and, in Gastric Cancer (GC), a large number of genes involved in carcinogenesis and clinical outcome accumulate aberrant methylation. Promoter methylation is an important mechanism of inactivation of tumour suppressor genes, in cancer cells.²

The process of promoter methylation is catalyzed by DNA methyltransferases (DNMTs) which organize, regulate and maintain properly mammalian genomes.³ Literature describes three types of DNMTs: DNMT1, over-expressed in human cancers including GC, that catalyze post-replication DNA methylation and maintain the methylation patterns during cell divisions^{4–6}; DNMT2, which is the most conserved and has a role in both DNA and RNA methylation⁷; and DNMT3, which is divided into two types (DNMT3a and DNMT3b) that are responsible for *de novo* methylation during gametogenesis and embryogenesis, and that also seem to be over-expressed in human cancers, including GC.^{3,6} Recently, literature has suggested that single nucleotide polymorphisms (SNPs) in DNMTs could promote functional consequences affecting methylation and therefore individual's susceptibility to cancer development.^{6,8}

Several studies have reported a potential association between SNPs in DNMTs and the GC susceptibility.^{3,9–11} Nevertheless, the observed associations of these studies are inconsistent and some studies are not large enough to take conclusions on the effect of DNMTs SNPs on GC. As a result, we have performed a systematic

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review and meta-analysis of all eligible studies to understand the possible association of SNPs in DNMTs genes and GC susceptibility.

Materials and methods

Search strategy

The present systematic review and meta-analysis followed PRISMA guidelines for Systematic Review and Meta-analyses. Literature was searched on PubMed data base until March 2016 applying the following query: “dna modification methylases”[MeSH Terms] OR (“dna”[All Fields] AND “modification”[All Fields] AND “methylases”[All Fields]) OR “dna modification methylases”[All Fields] OR (“dna”[All Fields] AND “methyltransferase”[All Fields]) OR “dna methyltransferase”[All Fields] OR DNMT[All Fields] AND (“polymorphism, genetic”[MeSH Terms] OR (“polymorphism”[All Fields] AND “genetic”[All Fields]) OR “genetic polymorphism”[All Fields] OR “polymorphism”[All Fields]) AND (“stomach neoplasms”[MeSH Terms] OR (“stomach”[All Fields] AND “neoplasms”[All Fields]) OR “stomach neoplasms”[All Fields] OR (“gastric”[All Fields] AND “cancer”[All Fields]) OR “gastric cancer”[All Fields]).

Inclusion and exclusion criteria

The literature search was limited to original studies performed in humans and no publication year restriction was applied. Only case-control studies with histologically-confirmed adenocarcinoma cases were included in this analysis. All DNMTs polymorphisms were selected for the analysis. Regarding the language of the papers, only studies written in English, Portuguese or Spanish were selected. Reviews, Meta-analysis, Systematic Reviews and studies not related to gastric cancer were excluded from this analysis.

Data extraction

Two authors performed the data extraction and all disagreements were resolved with the opinion of a third author. For each study, the following items were collected: country, type of study, ethnicity, matching criteria, age (years), polymorphisms, genotype method, and the number of cases and controls. Data concerning genotype distribution and the relative risk were also extracted for all polymorphisms included in the studies.

Statistical analysis

The evidence of Hardy Weinberg Equilibrium (HWE) in controls was recalculated in the present meta-analysis through the application of the online software (<http://www.had2know.com/academics/hardy-weinberg-equilibrium-calculator-2-alleles.html>). A p -value less than 0.05 of HWE was considered significant.

Meta-analysis for DNMTs SNPs was conducted by using the software Review Manager, version 5.3.5 (Copenhagen: The Nordic Cochrane Centre, The Cochrane Collaboration, 2014). The calculated OR (95% CI) was performed with Mantel–Haenszel statistical method and Random effects analysis model. A p -value less than 0.05 was considered significant.

Results

Literature search

A total of 35 papers were evaluated from which only 11 matched inclusion criteria. Seventeen articles were excluded due to the exclusion criteria by screening the titles and abstracts. After a

comprehensive evaluation of the remaining 18 articles, a total of 7 studies were excluded: 4 articles did not evaluate polymorphisms of DNMTs, only O6-methylguanine-DNA methyltransferase (MGMT)^{12–15}; 2 articles only reported mutations on DNMTs^{5,16}; 1 study was about allelic loss in gastric cancer.¹⁷ Finally, 11 studies involving a total of 3049 cases and 5185 controls were included in this analysis – Fig. 1.

Study characteristics

Table 1 describes the principal baseline characteristics of included studies. The majority of the studies were performed in China,^{3,8–11,18–21} one study in Iran²² and one in Japan.²³ Almost all studies have selected the controls and cases using age and gender as matching criteria, but some studies also used *Helicobacter pylori* infection status and Ethnicity. The number of cases and controls varies between the studies. A total of 24 polymorphisms were evaluated, 10 of DNMT1, 1 of DNMT2, 6 of DNMT3A and 7 of DNMT3B. The majority of studies were performed using Polymerase Chain Reaction–Restriction Fragment Length Polymorphism (PCR-RFLP) methodology, but other methods of genotyping were used like Taq-Man assay, High Resolution Melting (HRM) and Matrix Assisted Laser Desorption/Ionization – Time of Flight (MALDI-TOF). Supplementary Table 1 shows the characteristics of all polymorphisms included in the studies. The re-calculated HWE for controls is also shown in supplementary Table 2.

Genotyping results

The genotyping results of all studies are described in Table 2. A total of 10 SNPs in DNMT1 were studied by 3 different studies and only rs16999593, rs8101866 and rs2228611 have data from more than one population^{3,11,22} – Table 2.1. The risk analysis revealed that only one study reported a significant risk association (rs16999593) with GC development (OR=1.45; 95% CI=1.00–2.11; $p=0.05$, for C allele).

Only one study evaluated SNPs in DNMT2 and GC, revealing that rs112254413 A allele was associated with a protection for the development of GC (OR=0.15; 95% CI=0.08–0.27; $p<0.01$)³ – Table 2.2.

Thirteen SNPs of DNMT3 (6 from DNMT3A and 7 from DNMT3B) were evaluated by 9 different studies^{3,8–10,19–21,23,24} – Tables 2.3 and 2.4. The risk analysis have shown that DNMT3A rs7560488 (OR=1.65; 95%CI=1.19–2.29; $p=0.002$, for C allele) and rs36012910 (OR=2.36; 95%CI=1.35–4.14; $p=0.002$, for allele G) seem to be associated with an increased risk for GC.^{8,19} On the other hand, DNMT3A rs1550117 and rs13420867 show contradictory findings between the reported studies. In DNMT3B, only rs1569686 showed a statistically significant association ($p<0.01$) with GC, nevertheless the results are contradictory among the different studies.

Meta-analysis results

The meta-analysis was performed with data from all SNPs evaluated in more than one study – Figs. 2–4. A total of 6 SNPs were analyzed: DNMT1 rs16999593, rs8101866 and rs2228611; DNMT3A rs1550117 and rs13420827; and DNMT3B rs1569686. For the analysis, we have used the dominant genetic model, considering the risk genotype as the presence of any copy of the less frequent allele. We have also tested all data considering the Recessive Model for each SNP, and no significant change was observed in the reported data (data not shown). The majority of studies showed no significant heterogeneity in the data reported ($p>0.05$).

DNMT1: The analysis revealed no genetic heterogeneity ($p>0.05$) amongst the three SNPs tested. We found that only

Table 1
Characteristics of all studies included in the analysis.

Reference	Country (Ethnicity)	Matching Criteria	Cases		Controls		Gene	SNPS (rs)	Genotype method
			n	Age	n	Age			
Wang et al., 2015	China (Asian)	Age, Gender and <i>H. pylori</i>	447	NA	961	NA	DNMT3b	rs6119954, rs1569686, rs4911107, rs4911259, rs8118663	TaqMan assay
Wu et al., 2014	China (Asian)	Age and Gender	405	NA	408	NA	DNMT3a	rs7560488	High resolution melting
Cao et al., 2013	China (Asian)	Age, Gender and <i>H. pylori</i>	447	61.6	961	50.6	DNMT3a	rs1550117, rs13420827	TaqMan assay
Wu et al., 2012	China (Asian)	Age and Gender	340	62.2*	251	62.7*	DNMT3a	rs36012910	PCR-RFLP
Jiang et al., 2012	China (Asian)	Age, Gender and <i>H. pylori</i>	447	61.6	961	50.6	DNMT1	rs10420321, rs16999593, rs8101866, rs8111085, rs2288349	TaqMan assay
Yang et al., 2012	China (Asian)	Age and Gender	242	54.9 ± 12.5	294	58.4 ± 16.4	DNMT3a DNMT3b DNMT1 DNMT2	rs16999593, rs8101866, rs2228611, rs2114724, rs11254413, rs1550117, rs13420827, rs11887120, rs13428812, rs2424908	MALDI-TOF
Fan et al., 2010	China (Asian)	Age and Gender	208	65	346	71	DNMT3a	rs1550117	PCR-RFLP
Hu et al., 2010	China (Asian)	Age and Gender	259	64.0	262	65.1	DNMT3b	rs1569686, rs2424913	PCR-RFLP
Khatami et al., 2009	Iran (Turk and Fars)	Age, Gender and ethnicity	200	50 ± 13	200	58 ± 13	DNMT1	rs721186, rs13784, rs2228611, rs11488	PCR-RFLP
Aung et al., 2005	Japan (Asian)	NA	152	64.1*	247	43.9*	DNMT3b	rs2424913	PCR-RFLP
Wang et al., 2005	China (Asian)	Age and Gender	212	60.4	294	59.1	DNMT3b	rs2424913	PCR-RFLP

NA, not available; * Median (range); *H. pylori* – *Helicobacter pylori*; **DNMT1** – DNA (cytosine-5)-methyltransferase 1; **DNMT2** – TRNA (cytosine38-C5)-methyltransferase; **DNMT3a** – DNA (cytosine-5)-methyltransferase 3 alpha; **DNMT3b** – DNA (cytosine-5)-methyltransferase 3 beta; **MALDI-TOF** – Matrix-Assisted Laser Desorption/Ionization-Time of Flight; **PCR-RFLP** – Polymerase Chain Reaction-Restriction Fragment Length Polymorphism.

Table 2.1
Genotyping results for DNMT1 of all studies included.

Gene	SNP	Genotype distribution <i>n</i> (%)						Analysis model	OR (CI)	<i>p</i> value	Reference
		Cases			Controls						
DNMT1	rs10420321	AA 141 (31.5)	AG 203 (51.5)	GG 76 (17)	AA 328 (34.1)	AG 454 (47.2)	GG 179 (18.6)	CD	1.17 (0.88–1.55) (GG)	0.29	Jiang 2012
	rs16999593	TT 283 (63.3)	TC 144 (32.2)	CC 20 (4.5)	TT 659 (68.6)	TC 273 (28.4)	CC 29 (3)	CD	1.25(0.95–1.66) (CT)	0.18	Jiang 2012
		141 (58.3)	89 (36.8)	12 (5)	196 (66.7)	83 (28.2)	15 (5.1)	CD	1.47 (1.01–2.14) (TC)	0.11	Yang 2012
								Dominant	1.45 (1.00–2.11) (TC+ CC)	0.05	
	rs8101866	CC 238 (53.5)	CT 177 (39.8)	TT 30 (6.7)	CC 489 (50.9)	CT 402 (41.8)	TT 70 (7.3)	CD	0.81 (0.18–1.35) (CT)	0.11	Jiang 2012
		13 (5.4)	98 (40.1)	130 (53.9)	26 (8.8)	102 (34.7)	166 (56.5)	CD	1.20 (0.83–1.74) (TC)	0.17	Yang 2012
								Dominant	1.27 (0.89–1.82) (TC+ CC)	0.19	
	rs8111085	TT 143 (32)	TC 214 (47.9)	CC 90 (20.1)	TT 330 (34.3)	TC 447 (46.5)	CC 184 (19.1)	CD	1.18 (0.32–1.69) (CC)	0.38	Jiang 2012
	rs2288349	GG 249 (56.1)	GA 161 (36.3)	AA 34 (7.7)	GG 515 (53.6)	GA 372 (38.7)	AA 74 (7.7)	CD	0.81(0.50–1.33) (AA)	0.81	Jiang 2012
	rs2114724	CC 132 (54.5)	CT 97 (40.1)	TT 13 (5.4)	CC 162 (56.2)	CT 101 (35.1)	TT 25 (8.7)	CD	1.16 (0.81–1.68) (CT)	0.22	Yang 2012
	rs2228611							Dominant	1.23 (0.86–1.76) (CT+ TT)	0.27	
		GG 132 (54.5)	AG 97 (40.1)	AA 13 (5.4)	GG 160 (56.1)	AG 99 (34.7)	AA 26 (9.1)	CD	1.18 (0.81–1.71) (AG)	0.14	Yang 2012
								Dominant	1.26 (0.87–1.80) (AG+ GG)	0.22	
	rs721186	34 (34)	50 (50)	16 (16)	32 (32)	62 (62)	18 (18)	CD	1.13 (0.05–6.30) (AG)	NA	Khatami 2009
		CC 99 (99)	CT 1 (1)	TT –	CC 200 (100)	CT –	TT –	NA	NA	NA	Khatami 2009
rs13784	CC 200 (100)	CT –	TT –	CC 100 (100)	CT –	TT –	NA	NA	NA	Khatami 2009	
rs11488	AA 200 (100)	AT –	TT –	AA 200 (100)	AT –	TT –	NA	NA	NA	Khatami 2009	

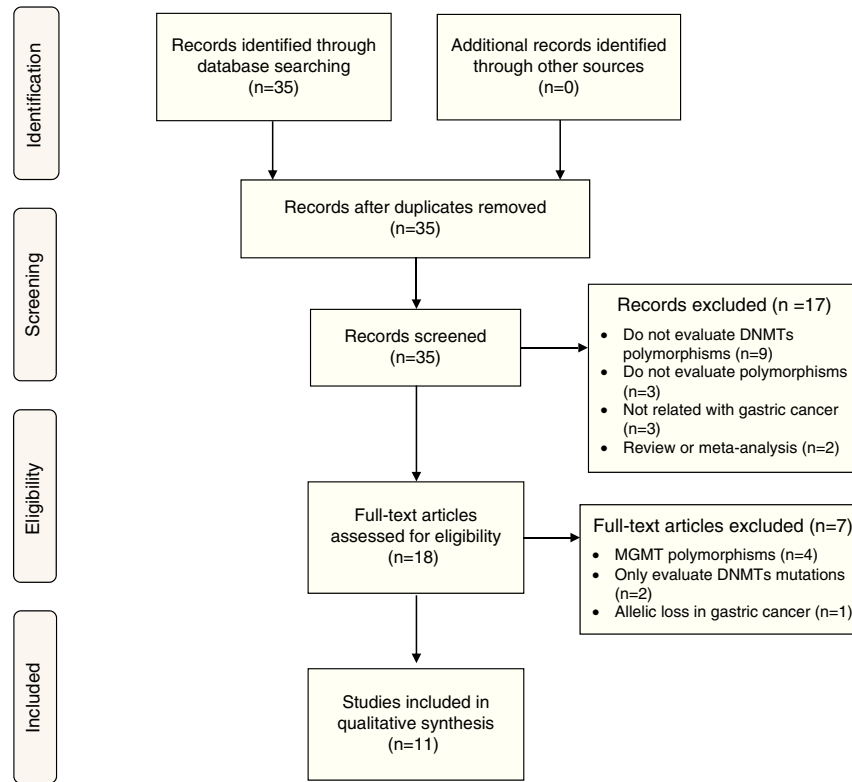


Fig. 1. Flow chart of studies identification, exclusion and inclusion. Adapted from: Moher D, Liberati A, Tetzlaff J, Altman DG, The PRISMA Group (2009). Preferred Reporting Items for Systematic Reviews and Meta-analyses: The PRISMA Statement. PLoS Med 6(6): e1000097. doi:10.1371/journal.pmed1000097.

Table 2.2
Genotyping results for DNMT2 of all studies included.

Gene	SNP	Genotype distribution n (%)						Analysis Model	OR (CI)	p value	Reference
		Cases			Controls						
DNMT2	rs11254413	GG	AG	AA	GG	AG	AA	CD Dominant	0.16 (0.09–0.28) (AG) 0.15 (0.08–0.27) (AG + AA)	<0.01 <0.01	Yang 2012
		204 (84.3)	15 (6.2)	23 (9.5)	187 (63.8)	91 (31.1)	15 (5.1)				

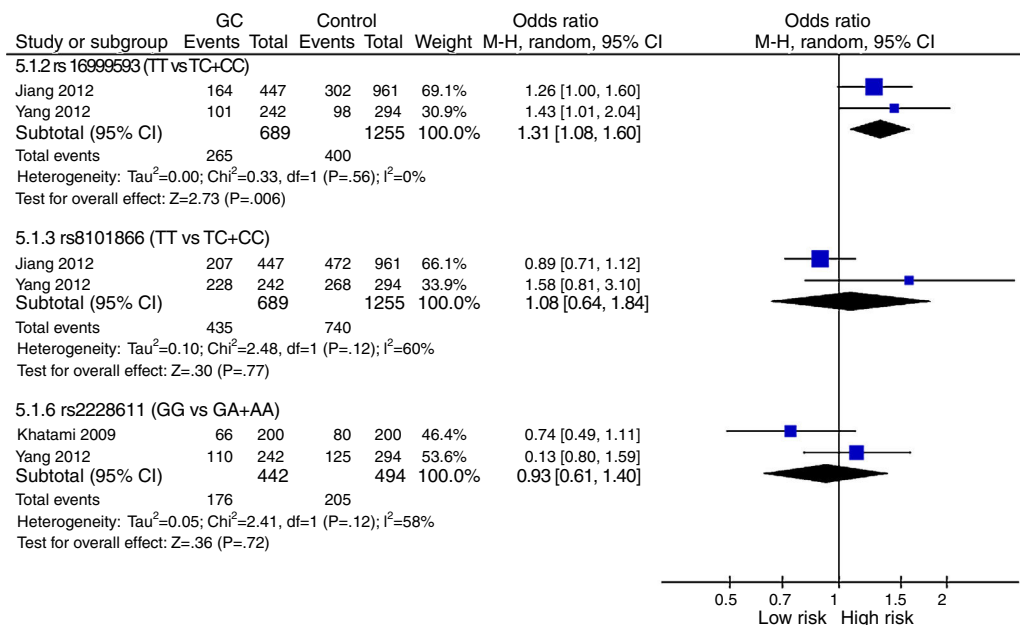


Fig. 2. Forest-plot for DNMT1 SNPs.

Table 2.3
Genotyping results for DNMT3A of all studies included.

Gene	SNP	Genotype distribution n (%)						Analysis model	OR (CI)	p value	Reference
		Cases			Controls						
DNMT3A	rs7560488	TT	TC	CC	TT	TC	CC	CD Dominant	1.73 (1.24–2.41) (TC); 2.50 (1.01–6.23) (CC) 1.65(1.19–2.29) (TC + CC)	0.01; 0.04 0.002	Wu 2014
		279 (68.9)	111 (27.4)	15 (3.7)	326 (79.9)	75 (18.4)	7 (1.7)				
	rs1550117	GG	GA	AA	GG	GA	AA	CD Dominant CD Dominant CD	1.10 (0.83–1.47) (AA) 1.06 (0.83–1.32) (GA + AA) 1.34 (0.55–3.29) (AA) 0.92 (0.63–1.34) (GA + AA) 6.63 (3.13–14.03) (AA)	0.51 0.51 0.74 0.68 0.00	Cao 2013 Yang 2012 Fan 2010
		289 (64.7)	142 (31.8)	16 (3.6)	640 (66.6)	288 (30)	33 (3.4)				
		157 (64.9)	74 (30.6)	11 (4.5)	191 (65)	93 (31.6)	10 (3.4)				
		102 (49)	75 (36.1)	31 (14.9)	218(63)	118 (34.1)	10 (2.9)				
	rs13420827	CC	CG	GG	CC	CG	GG	CD Dominant CD Dominant	1.06 (0.79–1.41) (GG) 1.05(0.83–1.33) (CG + GG) 0.68 (0.46–1.01) (CG) 0.66 (0.45–0.97) (CG + GG)	0.71 0.64 0.05 0.03	Cao 2013 Yang 2012
		295 (66)	133 (29.8)	19 (4.3)	618 (64.3)	304 (31.6)	39 (4.1)				
		167 (69)	61 (25.2)	14 (5.8)	183 (62.7)	99 (33.9)	10 (3.4)				
		AA	AG	GG	AA	AG	GG				
	rs36012910	AA	AG	GG	AA	AG	GG	CD Dominant	2.44 (1.37–4.33) (AG) 2.36 (1.35–4.14) (AG + GG)	0.002 0.002	Wu 2012
		288 (84.71)	51 (15)	1 (0.29)	234 (93.23)	17 (6.77)	0				
	rs11887120	TT	TC	CC	TT	TC	CC	CD	1.37 (0.88–2.13) (AA)	0.17	Wang 2015
		57 (23.6)	121 (50)	64 (26.4)	74 (25.3)	155 (53.1)	63 (21.6)				
rs13428812	AA	AG	GG	AA	AG	GG	CD Dominant	1.11 (0.58–2.12) (GG) 0.91 (0.64–1.31) (AG + GG)	0.84 0.63	Yang 2012	
	137 (56.6)	84 (34.7)	21 (8.7)	160 (55.4)	106 (36.7)	23 (8)					

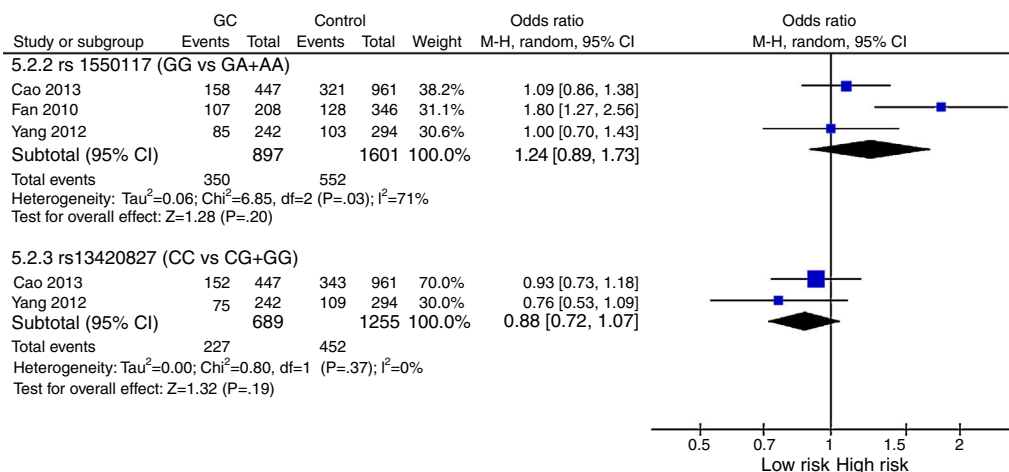


Fig. 3. Forest-plot for DNMT3A SNPs (Dominant model).

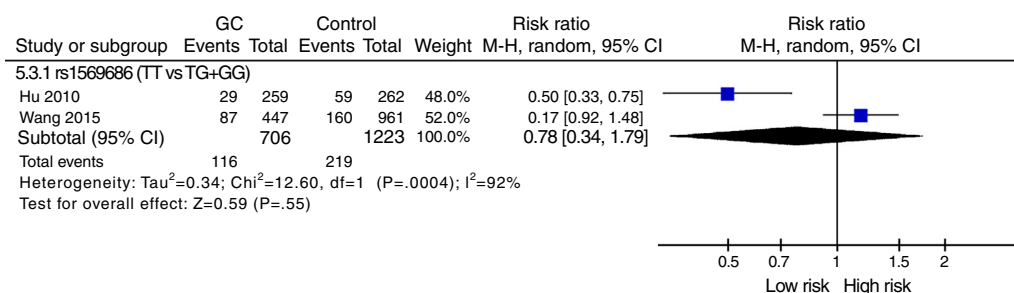


Fig. 4. Forest-plot for DNMT3B SNPs (Dominant model).

rs16999593 had a statistically significant result, revealing a 31% increased risk of GC development for TC + CC genotypes ($p = 0.006$; OR = 1.31; 95%CI = 1.08–1.60). The rs8101866 and rs2228011 showed no significant differences ($p > 0.05$) and no impact on the risk (OR = 1.08 and OR = 0.93, respectively) – Fig. 2.

DNMT3A: Statistical analysis showed heterogeneity of studies for rs1550117 ($p = 0.03$), and moreover no impact on the risk of GC development for either rs1550117 or rs13420827 ($p = 0.20$ and $p = 0.19$, respectively) – Fig. 3.

DNMT3B: One SNP (rs2424913) was excluded from the meta-analysis because there was no data for all genotypes in the three studies, making it impossible to estimate the OR value.^{9,23,24} The analysis also revealed that studies regarding rs1569686 are very contradictory, resulting in a high heterogeneity on studies ($p < 0.001$) and no association with GC – Fig. 4.

Discussion

Gastric carcinogenesis is a multistep process, where different factors are involved and epigenetic alterations seem to play an important role in early mechanisms of this process.⁶ DNA CpG methylation is catalyzed by DNMTs and modified DNA methylation has direct effect on the regulation of gene expression. Considering the significant impact of genetic susceptibility based on the genetic sequences of individuals, functional SNPs may affect DNMTs expression, and therefore, it is important to study SNPs in DNMTs and understand the effect that functional SNPs can have on protein activity.

Literature reveals that DNMTs are frequently overexpressed in GC.¹⁴ Recently, several studies have been performed to evaluate the impact of DNMTs polymorphisms in GC susceptibility.^{3,8–11,19–24} Hence, this study intends to resume the data published regarding the association of SNPs in DNMTs with GC. By searching literature, we have found 11 studies matching our inclusion criteria. One of

the first findings was that the majority of included studies were performed in Asiatic populations, where GC is extremely common, and therefore this is a major limitation since that is not possible to extrapolate the conclusions to others populations.²⁵

A total of 24 SNPs were found in this systematic review: 10 in DNMT1, 1 in DNMT2 and 13 in DNMT3. The heterogeneity found among the included studies is due not only to population differences but also to different genotyping methods. The genotyping methods used had different sensitivity/specificity: PCR-RFLP method has some limitations to genotype correctly the three genotypes, specially heterozygote genotypes²⁶; while TaqMan real-time PCR assays are more accurate and have less limitations and are a better candidate for SNP genotyping.²⁷ Furthermore, the number of cases and controls included are extremely different among studies and therefore more studies with bigger populations and more precise genotyping methods are still required.

Our results revealed that DNMT1 **rs16999593**, DNMT2 **rs11254413** and DNMT3A **rs7560488** and **rs36012910** are the only SNPs associated with GC.^{3,8,19} The DNMT1 **rs16999593** is characterized by a C>T variation, resulting in an Arg to His amino acid substitution at position 97 of the protein, leading to missense mutation that may affect the structure and function of DNMT1. Literature demonstrated that rs16999593 is significantly associated to different malignancies: Tao et al. showed that individuals with TC genotype had a 4-fold increased risk for sporadic triple-negative breast carcinoma²⁸; Xiang et al. also revealed that rs16999593 can provide protective effect for ductal breast carcinoma²⁹; and Yang et al. reported a 1.45-fold increased risk for TC + CC carriers and GC.³ Although these evidences, other studies do not show any association.^{11,30} DNMT2 **rs11254413** is characterized by a G>A substitution leading to an amino acid change, His to Tyr at position 101 of the protein that may affect DNMT2 function.³ The A-allele was associated with protection risk for GC by Yang et al., nevertheless, there is no functional study that support the biological role of this

Table 2.4
Genotyping results for DNMT3A of all studies included.

Gene	SNP	Genotype distribution n (%)						Analysis model	OR (CI)	p value	Reference
		Cases			Controls						
DNMT3B	rs6119954	GG 200 (44.8)	GA 199 (44.6)	AA 4 (10.6)	GG 433 (45.1)	GA 445 (46.3)	AA 83 (8.6)	CD	1.37 (0.88–2.13) (AA)	0.17	Wang 2015
	rs1569686	TT 360 (80.5)	TG 82 (18.3)	GG 5 (1.1)	TT 801 (83.4)	TG 150 (15.6)	GG 10 (1)	CD	1.33 (0.41–4.31) (GG)	0.64	Wang 2015
	rs4911107	AA 230 (88.8)	AG 27 (10.43)	GG 2 (0.77)	AA 203 (77.48)	AG 55 (20.99)	GG 4 (1.53)	Dominant	0.43 (0.26–0.72) (TG+GG)	<0.01	Hu 2010
	rs4911259	GG 360 (80.5)	GT 82 (18.3)	TT 5 (1.1)	GG 800 (83.2)	GT 151 (15.7)	TT 10 (1.1)	CD	1.08 (0.13–8.86) (GG)	0.94	Wang 2015
	rs8118663	AA 359 (80.3)	AG 83 (18.6)	GG 5 (1.1)	AA 799 (83.1)	AG 152 (15.8)	GG 10 (1.1)	CD	1.09 (0.13–8.88) (TT)	0.94	Wang 2015
	rs2424908	TT 126 (28.2)	TC 233 (52.1)	CC 88 (19.7)	TT 314 (32.7)	TC 473 (49.2)	CC 174 (18.1)	CD	1.32 (0.91–1.91) (GG)	0.15	Wang 2015
	rs2424913	CC 78 (32.2)	CT 114 (47.1)	TT 50 (20.7)	CC 99 (33.7)	CT 139 (47.3)	TT 56 (19.1)	CD	1.05 (0.64–1.71) (CC)	0.96	Yang 2012
			CC 257 (99.2)	CT 2 (0.8)	TT -	CC 259 (98.8)	CT 3 (1.2)	CD	1.49 (0.17–17.94) (CT)	>0.05	Hu 2010
			TT 152 (100)	-	-	TT 247 (100)	-	NA	NA	NA	Aung 2005
			TT 205 (96.7)	7 (3.3)	-	TT 279 (94.9)	15 (5.1)	Dominant	0.66 (0.26–1.67) (CT)	0.38	Wang 2005

NA, not available; CD, co-dominant; NR, no risk; DNMT1 – DNA (cytosine-5)-methyltransferase 1; DNMT2 – TRNA (cytosine38-C5)-methyltransferase; DNMT3a – DNA (cytosine-5)-methyltransferase 3 alpha; DNMT3b – DNA (cytosine-5)-methyltransferase 3 beta; SNP – Single Nucleotide Polymorphism; Allele: A – Adenine, C – Cytosine, G – Guanine, T – Thymine; OR (95% CI) – Odds Ratio (95% Confidence Interval).

SNP on DNMT2 function.³ The DNMT3A **rs7560488** (variation T>C) is a tagSNP that represents SNPs of the DNMT3A1 promoter with high linkage disequilibrium and has been associated with higher risk for GC in Asiatic population.⁸ Authors suggested that T to C change influences the binding of transcriptional factors and consequently expression levels of DNMT3A.⁸ The DNMT3A **rs36012910**, which is characterized by a A>G substitution, is located in the promoter region of DNMT3A and the substitution has been associated with DNMT3A altered activity.¹⁹ Wu et al. reveals that AG+GG genotypes were associated with a higher risk for GC.¹⁹

Regarding the data from the meta-analysis, we only were able to compare the results of 6 SNPs, which were described in more than one study. For DNMT1 analysis, 3 SNPs were studied (rs16999593, rs8101866 and rs2228611) and only **rs16999593** showed statistically significance^{3,22} – Fig. 2. The results showed an increased risk for TC+CC genotypes of GC development. Nevertheless, the low number of studies and the lack of studies from populations apart from China make it necessary to perform more studies with different populations to confirm the association of rs16999593 with GC. Concerning DNMT3A, 2 SNPs were included in the meta-analysis (rs1550117 and rs13420827) but no association with GC was found – Fig. 3. The analysis revealed heterogeneity in rs1550117 distribution in populations, especially when comparing the report by Fan et al. to the other studies (Fan et al., 2010). Nevertheless, even removing Fan et al. from the analysis there was no significant association of rs1550117 with GC development. Furthermore, the differences in genotype distribution might be explained because of different genotype methods used in the three studies – Table 1. Finally, we included one SNP in DNMT3B on the meta-analysis (rs1569686) that showed no association with GC – Fig. 4. The analysis revealed heterogeneity in the distribution and a significant difference in the OR of the two studies (OR=0.43 in Hu et al. and OR= 1.21 in Wang et al.).^{9,10}

Conclusions

We have found 11 articles studying SNPs in DNMTs but the variety of SNPs and the reduced number of studies per each SNP made this analysis complicated, allowing us to draw only some remarks and not strong conclusions. The data from studies revealed that rs11254413, rs7560488, rs36012910 and, specially, rs16999593 seem to be good candidates to be studied for their role in GC development. Nevertheless, our meta-analysis revealed that only one SNP (rs16999593) in DNMT1 was associated with GC development, especially TC+CC carriers.

More studies with different populations are required to prove if the SNPs selected in our analysis are associated with GC and furthermore, studies regarding biological importance of functional SNPs in DNMTs activity and expression are also needed to fully understand the impact of these SNPs in DNMTs on GC.

Conflicts of interest

The authors declare no conflicts of interest.

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at [doi:10.1016/j.pbj.2016.10.005](https://doi.org/10.1016/j.pbj.2016.10.005).

References

- Ziogas D, Roukos D. Epigenetics in gastric cancer: challenges for clinical implications. *Ann Surg Oncol.* 2009;16:2077–8. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/19408053> [cited 24.02.16].

- [2]. Feinberg AP, Vogelstein B. Hypomethylation distinguishes genes of some human cancers from their normal counterparts. *Nature*. 1983;301:89–92. <http://dx.doi.org/10.1038/3010890> [cited 24.02.16].
- [3]. Yang X-X, He X-Q, Li F-X, Wu Y-S, Gao Y, Li M. Risk-association of DNA methyltransferases polymorphisms with gastric cancer in the southern Chinese population. *Int J Mol Sci*. 2012;13:8364–78.
- [4]. Saito Y, Kanai Y, Sakamoto M, Saito H, Ishii H, Hirohashi S. Expression of mRNA for DNA methyltransferases and methyl-CpG-binding proteins and DNA methylation status on CpG islands and pericentromeric satellite regions during human hepatocarcinogenesis. *Hepatology*. 2001;33:561–8. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/11230735> [cited 25.02.16].
- [5]. Kanai Y, Ushijima S, Nakanishi Y, Sakamoto M, Hirohashi S. Mutation of the DNA methyltransferase (DNMT) 1 gene in human colorectal cancers. *Cancer Lett*. 2003;192:75–82. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/12637155> [cited 05.02.16].
- [6]. Yang J, Wei X, Wu Q, Xu Z, Gu D, Jin Y, et al. Clinical significance of the expression of DNA methyltransferase proteins in gastric cancer. *Mol Med Rep*. 2011;4:1139–43. Available from: <http://www.spandidos-publications.com/mmr/4/6/1139/Abstract> [cited 25.02.16].
- [7]. Schaefer M, Lyko F. Solving the Dnmt2 enigma. *Chromosoma*. 2010;119:35–40. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/19730874> [cited 14.02.16].
- [8]. Wu H, Zhang K, Gong P, Qiao F, Wang L, Cui H, et al. A novel functional TagSNP Rs7560488 in the DNMT3A1 promoter is associated with susceptibility to gastric cancer by modulating promoter activity. *PLOS ONE*. 2014;9:e92911.
- [9]. Hu J, Fan H, Liu D, Zhang S, Zhang F, Xu H. DNMT3B promoter polymorphism and risk of gastric cancer. *Dig Dis Sci*. 2010;55:1011–6.
- [10]. Wang C, Jia Z, Cao D, You L, Jin M, Wu X, et al. Polymorphism of DNA methyltransferase 3b and association with development and prognosis in gastric cancer. *PLoS ONE*. 2015;10:e0134059.
- [11]. Jiang J, Jia Z, Cao D, Jin M-S, Kong F, Suo J, et al. Polymorphisms of the DNA methyltransferase 1 associated with reduced risks of *Helicobacter pylori* infection and increased risks of gastric atrophy. *PLoS ONE*. 2012;7:e46058. Available from: <http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=3457938&tool=pmcentrez&rendertype=Abstract> [cited 19.04.16].
- [12]. Chikan NA, Bukhari S, Shabir N, Amin A, Shafi S, Qadri RA, et al. Atomic insight into the altered O6-methylguanine-DNA methyltransferase protein architecture in gastric cancer. *PLoS ONE*. 2015;10:e0127741. Available from: <http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=4444098&tool=pmcentrez&rendertype=Abstract> [cited 09.03.16].
- [13]. Imai Y, Oda H, Nakatsuru Y, Ishikawa T. A polymorphism at codon 160 of human O6-methylguanine-DNA methyltransferase gene in young patients with adult type cancers and functional assay. *Carcinogenesis*. 1995;16:2441–5. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/7586149> [cited 09.03.16].
- [14]. Lin J, Zeng RM, Li RN, Cao WH. Aberrant DNA methylation of the P16, MGMT, and hMLH1 genes in combination with the methylenetetrahydrofolate reductase C677T genetic polymorphism and folate intake in gastric cancer. *Genet Mol Res*. 2014;13:2060–8.
- [15]. Xiong H-L, Liu X-Q, Sun A-H, He Y, Li J, Xia Y. Aberrant DNA methylation of P16, MGMT, hMLH1 and hMSH2 genes in combination with the MTHFR C677T genetic polymorphism in gastric cancer. *Asian Pac J Cancer Prev*. 2013;14:3139–42.
- [16]. Oh HR, An CH, Yoo NJ, Lee SH. Somatic mutations of amino acid metabolism-related genes in gastric and colorectal cancers and their regional heterogeneity – a short report. *Cell Oncol (Dordr)*. 2014;37:455–61. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/25450519> [cited 09.03.16].
- [17]. Kim MS, Kim YR, Yoo NJ, Lee SH. Mutational analysis of DNMT3A gene in acute leukemias and common solid cancers. *APMIS*. 2013;121:85–94. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/23031157> [cited 03.03.16].
- [18]. Wang Y, Lou B, Yan LP, Huang BH, Zhao P. Relationship between Epstein-Barr virus-encoded proteins with cell proliferation, apoptosis, and apoptosis-related proteins in gastric carcinoma. *World J Gastroenterol*. 2005;11:3234–9.
- [19]. Wu Q, Lu S, Wang L, Hu J, Qiao F, Qiu X, et al. DNMT3A rs36012910 A>G polymorphism and gastric cancer susceptibility in a Chinese population. *Mol Biol Rep*. 2012;39:10949–55.
- [20]. Cao X-Y, Jia Z-F, Cao D-H, Kong F, Jin M-S, Suo J, et al. DNMT3a rs1550117 polymorphism association with increased risk of *Helicobacter pylori* infection. *Asian Pac J Cancer Prev*. 2013;14:5713–8.
- [21]. Fan H, Liu D, Qiu X, Qiao F, Wu Q, Su X, et al. A functional polymorphism in the DNA methyltransferase-3A promoter modifies the susceptibility in gastric cancer but not in esophageal carcinoma. *BMC Med*. 2010;8:12.
- [22]. Khatami F, Noorinayer B, Ghiasi S, Mohebi R, Hashemi M, Zali MR. Lack of effects of single nucleotide polymorphisms of the DNA methyltransferase 1 gene on gastric cancer in Iranian patients: a case control study. *Asian Pac J Cancer Prev*. 2009;10:1177–82.
- [23]. Aung PP, Matsumura S, Kuraoka K, Kunimitsu K, Yoshida K, Matsusaki K, et al. No evidence of correlation between the single nucleotide polymorphism of DNMT3B promoter and gastric cancer risk in a Japanese population. *Oncol Rep*. 2005;14:1151–4.
- [24]. Wang Y-M, Wang R, Wen D-G, Li Y, Guo W, Wang N, et al. Single nucleotide polymorphism in DNA methyltransferase 3B promoter and its association with gastric cardiac adenocarcinoma in North China. *World J Gastroenterol*. 2005;11:3623–7.
- [25]. Karimi P, Islami F, Anandasabapathy S, Freedman ND, Kamangar F. Gastric cancer: descriptive epidemiology, risk factors, screening, and prevention. *Am Assoc Cancer Res*. 2014;23:700–13. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/24618998> [cited 26.07.16].
- [26]. Davis AHT, Wang J, Tsang TC, Harris DT. Direct sequencing is more accurate and feasible in detecting single nucleotide polymorphisms than RFLP: using human vascular endothelial growth factor gene as a model. *Biol Res Nurs*. 2007;9:170–8. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/17909169> [cited 19.04.16].
- [27]. Shen G-Q, Abdullah KG, Wang QK. The TaqMan method for SNP genotyping. *Methods Mol Biol*. 2009;578:293–306. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/19768602> [cited 10.02.16].
- [28]. Tao R, Chen Z, Wu P, Liu C, Peng Y, Zhao W, et al. The possible role of EZH2 and DNMT1 polymorphisms in sporadic triple-negative breast carcinoma in southern Chinese females. *Tumour Biol*. 2015;36:9849–55. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/26162541> [cited 19.04.16].
- [29]. Xiang G, Zhenkun F, Shuang C, Jie Z, Hua Z, Wei J, et al. Association of DNMT1 gene polymorphisms in exons with sporadic infiltrating ductal breast carcinoma among Chinese Han women in the Heilongjiang Province. *Clin Breast Cancer*. 2010;10:373–7. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/20920981> [cited 19.04.16].
- [30]. Ye C, Beeghly-Fadiel A, Lu W, Long J, Shu XO, Gao Y-T, et al. Two-stage case-control study of DNMT-1 and DNMT-3B gene variants and breast cancer risk. *Breast Cancer Res Treat*. 2010;121:765–9. Available from: <http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=3493111&tool=pmcentrez&rendertype=Abstract> [cited 19.04.16].